[1:3] Site-differentiated and sulfide-bridged cubane clusters in chemistry and biology

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Abstract: A subset of protein-bound Fe₄S₄ cubane-type clusters occurs in a [1:3] iron site-differentiated form in which the terminal ligand at the unique site is not a cysteinyl group. Reactions of synthetic site-differentiated [Fe₄S₄]²⁺ clusters are summarized, including removal of the unique iron atom to afford the [Fe₃S₄]⁰ cuboidal cluster, thereby completing the synthesis of analogues of native iron-sulfur clusters with nuclearity ≤4. The cuboidal cluster supports metal ion insertion reactions leading to a set of heterometal cubane type clusters MFe₃S₄, whose redox potentials have been determined as a function of M. Sulfide-bridged double cubanes [MFe₃S₄]–S–[MFe₃S₄] (M = Mo, Fe; M_2 = MoFe), another type of site-differentiated cluster, have been prepared as precursors in potential cluster rearrangement reactions leading to the core [MFe₆S₉] with the same topology as the FeMo-cofactor of nitrogenase.

INTRODUCTION

The pervasiveness in biology of iron-sulfur clusters containing the rhomboidal $Fe_2(\mu_2-S)_2$, cuboidal $Fe_3(\mu_2-S)_2$ S)₃(μ_3 -S) and cubane-type Fe₄(μ_3 -S)₄ cores is well recognized (1,2). Over a ca. 25-year period, cluster structures have been established by protein crystallography (3), physicochemical properties elucidated in considerable detail, and analogue clusters synthesized. These clusters are conventionally bound to proteins entirely through S·Cys ligands, in which form they have an electron storage and transfer function. Of particular current interest are Fe₄S₄ clusters that are rendered [1:3] site-differentiated by virtue of non-cysteinyl coordination at one iron site. The chemistry of these clusters is considerably expanded beyond electron transfer by this departure from standard terminal ligation. Native [1:3] site-differentiated clusters 1-4 are set out in Fig. 1. Cluster 1 is the catalytic center in aconitase and certain other hydro-lyases (4). Substrate is bound, activated, and transformed at the unique iron site. Under our previous definition, sites 2 and 3 are biological bridged assemblies (5). Assembly 2 represents the two different catalytic centers of carbon monoxide dehydrogenase (Clostridium thermoaceticum) (6), center A for acetyl coenzyme A synthase activity and center C for CO/CO₂ interconversion. In each, a nickel atom with a coordination sphere of incompletely defined geometry and ligation is bridged through an unknown atom or group X to one site of an Fe₄S₄ cluster.

Fig. 1. Native [1:3] Site-Differentiated Fe₄S₄ Clusters

Assembly 3 is the active site of sulfite reductase (*Escherichia coli*) (7), wherein a siroheme group is coupled to an Fe_4S_4 cluster through its unique iron site by means of an $Fe_-(\mu_2-S\cdot Cys)$ —Fe bridge containing one very long bond. Substrate is bound in an axial position trans to the bridge, and electrons are presumably injected one at time into the cluster and flow to siroheme and then to substrate. Cluster 4 is the distal component in the linear cluster arrangement $Fe_4S_4|Fe_3S_4|Fe_4S_4$, which is apparently the conduit for electron flow in the uptake [NiFe]-hydrogenase of *Desulfovibrio gigas* (8). This is the only proven example of imidazole ligation to a native Fe_4S_4 cluster. These and other observations over the last decade, including formation of protein-bound Fe_3S_4 clusters from [1:3] clusters (9), have made imperative the development of site-differentiating ligands that facilitate an extensive examination of the chemistry of such clusters.

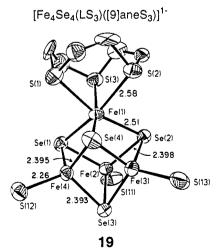
SYNTHETIC [1:3] SITE-DIFFERENTIATED Fe₄Q₄ CLUSTERS

Two types of site-differentiating ligands have been developed in this laboratory (10-12). The more successful is the semirigid trithiolate cavitand ligand LS₃, which is configured to bind efficiently cubane-type Fe₄Q₄ cores as the clusters $[Fe_4Q_4(LS_3)L']^{2^2}$ (5). The indicated mode of cluster binding has been proven

crystallographically (11, 13). In the ababab conformation shown, three coordinating arms (a) are buttressed on the same side of the central benzene ring by the three p-tolylthio legs (b). Regiospecific substitution reactions occur at the unique iron site. About 100 clusters with various ligands L' (Q = S, Se) have been generated in solution and/or isolated; without exception, their ¹H NMR spectra indicate trigonal symmetry in solution. The clusters 5 have been applied to a number of problems: (i) synthesis of a large set of substituted clusters (11,13-16), including factors such as stabilization of five- and six-coordinate unique sites with attendant modulation of redox potentials, charge distribution, and cluster spin state (15,16); (ii) formation of bridged double cubanes with independent and coupled electron transfer reactions across bridges of variable length (14,17,18); (iii) synthesis of sulfide-bridged analogues of assembly 3 (19); (iv) elucidation of comparative isotropic shifts, redox potentials, and ligand binding affinities of $[Fe_4S_4]^{2+}$ and $[Fe_4S_4]^{2+}$ clusters (13); (v) preparation of cuboidal

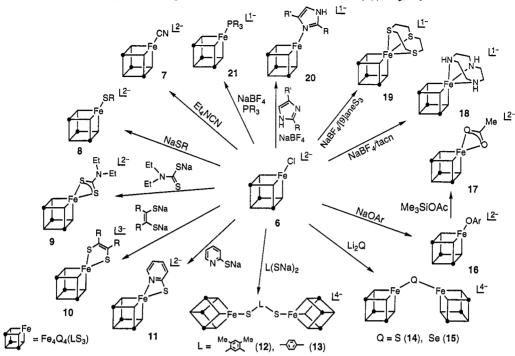
 $[\text{Fe}_3\text{S}_4(\text{LS}_3)]^3$ by iron atom removal (20); (vi) formation of heterometal cubane clusters by metal ion insertion into a cuboidal cluster (21). Applications (i)-(iv) have been made possible largely by the hypersensitivity of the isotropically shifted ^1H resonances of the $[\text{Fe}_4\text{Q}_4]^{2+}$ clusters 5 to the nature of L'.

The isotropic components of the chemical shifts of the 4-Me, 5-H, and 6-Me substituents of **5** in the $[Fe_4Q_4]^{2+}$ oxidation state are mainly or exclusively contact in origin, and for this reason are responsive to changes in the ligand L'. Figure 2 represents a partial summary of the substitution reactions at the unique iron site (10,11, 13-20) monitored by ${}^{1}H$ NMR. Although others may be used, $[Fe_4Q_4(LS_3)Cl]^{2-}$ (**6**) are convenient starting clusters owing to the lability of chloride. Clusters with anionic ligands (**7-17**), including those with 5-



coordinate sites (9-11) and bridges between two clusters (12-15), are readily formed. Neutral ligands can be introduced, as in clusters 18-21 which include examples of six-coordinate sites (18, 19). A number of useful results follow from the substituted clusters, of which two related to application (i) are noted here. The structure of $[Fe_4Se_4(LS_3)([9]aneS_3)]^{1-}$ (19) proves that an iron site in a $[Fe_4Q_4]^{2+}$ cluster can stabilize six-coordination with nonoxygen ligands. In the enzyme-inhibitor complexes of aconitase (4b), five- and six-coordinate sites FeS₃O_{2,3} have been shown crystallographically. An estimate of the intrinsic effect of imidazole ligation on the [Fe₄S₄]^{2+/1+} redox potential can be obtained from clusters 8 and 20. In acetonitrile, the difference in potential of the reference cluster [Fe₄S₄(LS₃)(SEt)]²⁻ (8) and $[Fe_4S_4(LS_3)(4-MeIm)]^{1-}$ (20, R' = Me) is -330 mV; i.e., the latter is more easily reduced as would be expected from is lesser negative charge. However, the potential difference between the

Fig. 2. Site-Specific Substitution Reactions of [Fe₄Q₄(LS₃)Cl]²-



reference $[Fe_4S_4(S\cdot Cys)_4]$ cluster proximal to the active site in *D. gigas* hydrogenase and distal $[Fe_4S_4(S\cdot Cys)_3(N\cdot His)]$ (4) is only -60 mV at pH 8 (22). While the sign of the difference is the same as with 8 and 20, evidently there are protein environmental and pH effects that significantly attenuate the potential difference between the proximal and distal clusters compared to the estimated intrinsic value. Applications (ii), (v), and (vi) and double cubane clusters are considered separately below.

CUBOIDAL Fe₃S₄ CLUSTERS

Synthesis. The cuboidal cluster $[Fe_3S_4(S\cdot Cys)_3]$ is now commonly encountered in iron-sulfur proteins (1,9), and is recognized by characteristic EPR, MCD, and Mössbauer spectroscopic properties of the $[Fe_3S_4]^{1+,0}$ oxidation states. However, the obtainment of a synthetic analogue has proven to be more difficult than for Fe_2S_2 and Fe_4S_4 clusters, which can be readily prepared by self-assembly methods. The problem was solved by using the reaction scheme in Figure 3 starting with site-differentiated cluster 8 (20). Labilization of the unique site by thiolate substitution followed by triflate displacement with the *fac*-tridentate ligand N-methylimidodiacetate (Meida) affords 23. Reaction of the latter with excess Meida gives the desired cluster $[Fe_3S_4(LS_3)]^{3-}$ (24) in 76% yield based on 8. All steps were performed in acetonitrile; the selenide cluster was prepared by an analogous reaction scheme. The cluster can also be synthesized by the reaction of the cubane-type cluster $[(OC)_3MoFe_3S_4(LS_3)]^{3-}$ with CO; the molybdenum fragment is removed as $[Mo(CO)_{6-n}(MeCN)_n]$ (n = 3-6) (23). The cluster core approaches idealized trigonal symmetry. The collective geometrical and

SEt
$$OSO_2CF_3$$
 OSO_2CF_3 OSO_2CF_3

electronic structural data demonstrate that 24 is an accurate analogue of the protein-bound cluster. As such, it demonstrates several important points. Protein structure itself is not necessary to stabilize cuboidal geometry. However, the semirigid LS₃ ligand substantially enhances stability inasmuch as its replacement with simple monothiolate ligands results in relatively rapid cluster decay (20). The S=2 ground state of protein-bound $[Fe_3S_4]^0$ clusters, arising from antiparallel coupling of the S=9/2 spin from double exchange within an $Fe^{3+}Fe^{2+}$ pair and S=5/2 of an Fe^{3+} site, and attendant Mössbauer and magnetic parameters are reproduced almost exactly by 24 (20). Therefore, this electronic state is an intrinsic property of cuboidal $[Fe_3S_4]^0$, not set by protein structure.

Metal Ion Incorporation. While the cuboidal Fe_3S_4 cluster has been detected in over 30 proteins, its biological function other than electron transfer remains unclear. The best recognized non-redox reaction of protein-bound clusters is metal-ion incorporation. Incubation of a protein with excess metal salt followed by chromatographic purification affords MFe_3S_4 clusters, which may be oxidized or reduced in subsequent reactions (24,25). Their cubane-type structures have been established by spectroscopy and comparison with synthetic clusters of known structure (26). Thus far, clusters with the cores $[MFe_3S_4]^{2+.1+}$ (M = Mn, Co, Ni, Cu, Zn, Cd, Tl) have been identified in proteins. Isolation of 24 permits the investigation of incorporation reactions under near-stoichiometric conditions in synthetic systems (21). Leading results are summarized in Fig. 4, where it is shown that 24 binds $M^{2+} = Fe$ (6), Co (25), Ni (26), $M^{1+} = Cu$ (27,28), Tl (30), Ag (31,32), and $M^0 = Mo$, W (29), and is, therefore, a cluster ligand. Clusters are isolated in the $[MFe_3S_4]^0$ (29) and $[MFe_3S_4]^{1+}$ (all others) oxidation states. Spectroscopic results have shown that except for 25 and 26, with the $[Fe_3S_4]^{1-}$ (S = 5/2) fragment bound to M^{2+} , the clusters contain $[Fe_3S_4]^0$ (S = 2) coordinated to $M^{1+,0}$. Redox reactions largely change the electron distribution in the Fe_3S_4 portions of the clusters. The dinegative clusters in Fig. 4 permit determination of the heterometal dependence of redox potential for the couple $[MFe_3S_4]^{1+,0}$ at constant cluster charge and terminal ligands. For 25-27, 30 (M ligand absent), and 32 in acetonitrile, the

Fig. 4. Formation of Heterometal Clusters [MFe₃S₄]

interval of potentials is ca. 400 mV and the order is Co < Ni < Cu \leq Ag < Tl (21). We take this to be the intrinsic order. Progressions of protein-bound cluster potentials are not invariant among proteins and do not tend to follow this order (24); however, terminal ligands at the M sites are unestablished and protein environmental effects are unknown. Because the potential of the couple [24]^{2-/3-} ([Fe₃S₄]^{1+/0}) is -0.79 V in acetonitrile vs. SCE, incorporated heterometal ions afford clusters whose potentials (Fig. 4) for the same

cluster and core charge couples are some 400 to 800 mV lower than that of the free cuboidal cluster. The $[MFe_3S_4]^0$ oxidation state apparently requires potentials too negative for stabilization in proteins in aqueous solution. Metal ion incorporation is a new reaction category for synthetic iron-sulfur clusters. With the exception of highly thiophilic Cu^{1+} and Tl^{1+} in the cores $[MFe_3S_4]^{2+}$, heterometal binding requires the $[Fe_3S_4]^{0.1-}$ oxidation states, in which the μ_2 -S atoms are more nucleophilic. All earlier MFe_3S_4 clusters were synthesized by self-assembly reactions or reductive rearrangement of linear Fe_3S_4 clusters $(M^{1+0} + [Fe_3S_4]^{1+} \rightarrow [MFe_3S_4]^{2+,1+}$ (26)). The possible natural occurrence of MFe_3S_4 clusters is a matter of continuing interest.

BRIDGED CLUSTER ASSEMBLIES

Nitrogenase Clusters. Lest one consider that the challenge of understanding native iron-sulfur clusters at a rational level of detail has largely been met, attention is directed to the metal clusters of nitrogenase (27), the P-cluster 33 and the cofactor cluster 34 (M or FeMo-co) whose cores have the composition Fe₈S₇ and MoFe₇S₉, respectively. The seminal contributions by Rees and coworkers (28) in the xray structure determination of nitrogenase proteins has reinvigorated the field of iron-sulfur chemistry and biochemistry, and introduced a new level of complexity and sophistication into the subject of natural and synthetic ironsulfur clusters. The complexity derives from the structures themselves, originating only in a protein venue and manifesting the highest nuclearity of any protein-bound metal-sulfur cluster, and imposes upon the ultimate disclosure of cluster formation, properties, and function a level of sophistication exceeding that required for the smaller clusters. Cluster 33 in the POX paramagnetic oxidation state (not shown) consists of an Fe₄S₄ cluster with one μ_4 -S atom bound to an iron atom of an Fe₃S₄ cuboidal fragment. In the more reduced PN state, this atom becomes at least μ_5 , interacting with two iron atoms of the cuboidal fragment at 2.4 Å; a third iron atom (dashed line) is located

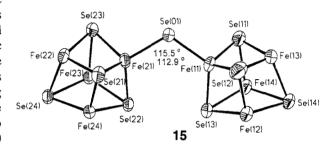
at 2.9 Å. In both states, the two cluster fragments are bridged by two Cys·S ligands. (These structures supplant an earlier version in which two $F_{4}S_{4}$ clusters were connected by a S–S bond (27)). In passing from P^{OX} to P^{N} , several changes in terminal ligation occur concomitant with an increase in interfragment Fe–S bonds (28). Cluster 34 is composed of $F_{4}S_{3}$ and $MoF_{3}S_{3}$ cuboidal fragments bridged by three μ_{2} -S atoms which are bound to three-coordinate iron atoms. This structure is maintained in the diamagnetic M^{OX} and the paramagnetic M^{N} (S = 3/2) states. The P-cluster apparently is the ultimate electron donor to FeMo-co, which is the catalytic site for dinitrogen reduction. Because FeMo-co can be removed with core intact from the protein by disruption of the Fe–S·Cys and Mo–N·His interactions in 34, it has long been recognized as a potentially viable object of synthesis. When not ensconced in a protein environment that controls gated electron and proton flux to the site of catalysis, the cofactor itself is not expected to effect ammonia formation from dinitrogen. However, its preparation in substantial yield from non-protein components must involve new concepts and reactions in cluster synthesis, and the study of accessible oxidation states and their interactions with substrates should be most informative.

<u>Sulfide-Bridged Double Cubanes</u>. One conceptual route to clusters with the topology of **34** begins with this largely uninvestigated cluster type. The first evidence that Fe_4S_4 clusters could be linked occurred in the form of the double cubane clusters **12-14**, which are derived from site-differentiated **6** by chloride displacement (Fig. 2) (17). Shortly thereafter, it was shown that undifferentiated $[Fe_4S_4Cl_4]^2$ reacts with stoichiometric Li_2S to form $\{[Fe_4S_4Cl_3]_2S\}^4$, whose bridged double cubane structure was established by an x-ray structure determination (29). Note that the $[Fe_4S_4]-S-[Fe_4S_4]$ core of double cubanes has the same sulfur-metal ratio as $M_2Fe_6S_9$, where $M=M_0+Fe$ in **34** and Mo or Fe in possible symmetrized synthetic clusters. Removal of suitably labile ligands (chloride, aryloxide) by chemical reaction or dissociation (e.g., triflate) followed by

core rearrangement could yield the FeMo-co topology, as in the generalized core reaction [MFe₃(μ_3 -S)₄]₂(μ_2 -S) \rightarrow [MFe₃(μ_3 -S)₃]₂(μ_2 -S)₃. To proceed with this concept (18), it was first necessary to develop a synthetic scheme for double cubanes and means of structure proof without complete reliance on x-ray crystallography, in the event that such compounds could not always be crystallized.

The heterogenous reactions $[Fe_4Q_4(LS_3)Cl]^{2^2} + Li_2Q$ (or $Na_2S) \rightarrow \{[Fe_4Q_4(LS_3)]_2Q\}^{4^2}$ (14,15) in acetonitrile afford ca. 90% yields of the product clusters. The double cubane structure of 15 was proved crystallographically (18). In this case, the asymmetric unit contained two cluster anions with the indicated

bridge angles, leading to intercubane Fe···Fe separations of 3.85 Å and 3.79 Å. This cluster, **14**, and {[Fe₄S₄Cl₃]₂S}⁴ proved useful for developing two non-crystallographic criteria for sulfide-bridged double cubane formation (18). (i) Cluster parent ions (associated with R₄N⁺ counterions), having characteristic isotope patterns, are observable by electrospray mass spectrometry. (ii) Two distinct reduction steps, separated by 250-310 mV in acetonitrile, are found in voltammetry.



Related single cubanes show one such step, always at less negative potentials, indicating the stepwise reduction of individual clusters ($[Fe_4S_4]^{2+/1+}$) in intact double cubanes. These criteria are also expected to apply to heterometal sulfide-bridged double cubanes, and closely related species. Among the latter is $[VFe_4S_6(PEt_3)_4]_2S$, derived from the cuboidal cluster $[VFe_4S_6(PEt_3)_4Cl]$ (30), which meets these criteria and in addition shows two discrete oxidation steps (18).

Synthesis of sulfide-bridged heterometal double cubanes is somewhat more involved because it requires the preparation of appropriate single-cubane precursors. The latter are, however, members of a widely explored group of heterometal MFe₃S₄ cubanes (9). One reaction scheme that has been developed is set out in Fig. 5 (18). The cluster $[(C_6Cl_4O_2)(\text{solv})\text{MoFe}_3S_4Cl_3]^{2-}$ (35, cat = catecholate(2-)), available in three steps from a

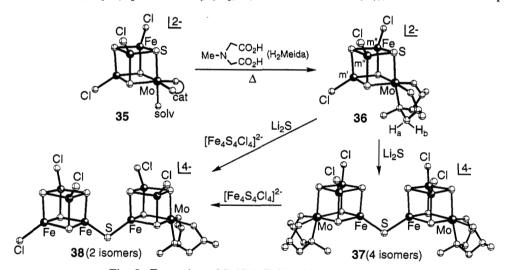


Fig. 5. Formation of Sulfide-Bridged Double Cubanes

self-assembled bridged double cubane (31), is converted by site-specific ligand substitution to $[(Meida)MoFe_3S_4Cl_3]^2$ (36) (18,32), containing a tight-binding tridentate ligand to prevent bridge formation at the Mo site. This cluster has mirror symmetry, two types of iron sites (m',m"), and diastereotopic methylene protons ($H_{a,b}$). Coupling of 36 with Li_2S affords the symmetrical double cubane 37 which, owing to different relative arrangements of the two Meida ligands, exists as four geometrical isomers. This compound was isolated as $(Et_4N)_4[37]$ -2LiCl in 64% yield from reaction 1 in acetonitrile. The presence of isomers is indicated by multiple isotropically shifted $H_{a,b}$ signals and ¹⁹F resonances of the substitution product of 37,

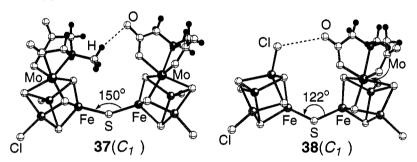
{[(Meida)MoFe₃S₄(S-p-C₆H₄F)₂]₂S}⁴⁻. Despite multiple attempts, it has not proven possible to obtain diffraction-quality crystals of any compound containing 37. However, cluster 37 meets criteria (i) and (ii) for a sulfide-bridged structure; in DMF two reduction steps are observed for 37 (-1.01, -1.34 V vs. SCE) compared to one step for 36 (-0.81 V). In addition to the formation of 37, it is also possible to generate unsymmetrical sulfide-bridged double cubane 38 in two systems, both demonstrated to be reversible equilibria in Me₂SO. Cluster 38 can exist as two isomers, depending on whether the bridge is formed at an m' or m" site. When the solid isolated from reaction 2 was redissolved in Me₂SO, the product mol ratio 38:37:36 \approx 9:3:1. Equimolar reaction 3 was also shown to generate 38 together with 36. Double cubane 38 was positively

$$2[(Meida)MoFe3S4Cl3]2- + Li2S \rightarrow \{[(Meida)MoFe3S4Cl2]2S\}4- + 2LiCl$$
 (1)

$$3[(Meida)MoFe_{3}S_{4}Cl_{3}]^{2-} + 3[Fe_{4}S_{4}Cl_{4}]^{2-} + 3Li_{2}S \neq \{[(Meida)MoFe_{3}S_{4}Cl_{2}]S(Fe_{4}S_{4}Cl_{3})\}^{4-} + \{[(Meida)MoFe_{3}S_{4}Cl_{2}]_{2}S\}^{4-} + [(Fe_{4}S_{4}Cl_{3})_{2}S]^{4-} + 6LiCl$$
 (2)

$$\{[(Meida)MoFe_{3}S_{4}Cl_{2}]_{2}S\}^{4-} + [Fe_{4}S_{4}Cl_{4}]^{2-} \neq \{[(Meida)MoFe_{3}S_{4}Cl_{2}]S(Fe_{4}S_{4}Cl_{3})\}^{4-} + [(Meida)MoFe_{3}S_{4}Cl_{3}]^{2-}$$
(3)

identified by mass spectrometry. When the solid from reaction 2 was treated with acetonitrile, only 38 was detected by ¹H NMR owing to the low solubility of $(Et_4N)_2[37]$. (Any $[(Fe_4S_4Cl_3)_2S]^{4-}$ present cannot be detected in this way.) These results suggest the stability order 38 > 37, with the probable origin being differing steric repulsions across the bridge. There exist conformations of 37 in which the two Meida moieties come into steric contact, whereas in 38 Meida/Cl⁻ interactions are less destabilizing. Conformations of these two clusters have been analyzed, and a stability order including all isomers of each has been deduced (18). In the most crowded conformations, the bridge angle in $38(C_1)$ much more closely approaches the unstrained value of 102° in $\{[Fe_4S_4Cl_3]_2S\}^{4-}$ (29) than does the angle in $37(C_1)$, indicating less steric destabilization.



Alternative Approaches to Biological Analogues. By the foregoing means, clusters containing the bridged cores $[MoFe_3S_4-S-MoFe_3S_4]^{4+}$, $[MoFe_3S_4-S-Fe_4S_4]^{3+}$, and $[Fe_4S_4-S-Fe_4S_4]^{2+}$ have been prepared. The compositions of the first and third of these approach that of FeMo-co (34, MoFe₇S₉), and the second is identical; 38 is the first synthetic cluster with this property. These clusters or suitably substituted derivatives serve as viable candidates for the foregoing cluster rearrangement reaction potentially affording one or more of the cores $[M_2Fe_6S_9]^{2+}$ (M = Mo, Fe; M_2 = Mo + Fe). It remains to be seen whether the unique and complex structure of FeMo-co can be created without benefit of proteins, its stability outside a protein environment notwithstanding. Current biosynthetic evidence indicates that FeMo-co is assembled on gene products and transferred intact to the apo-MoFe protein of nitrogenase (33). The proteins involved presumably act as scaffolds, on which portions of the cofactor are formed and ultimately combined. Indeed, the problem of nonbiological FeMo-co synthesis raises a broader issue. Can meaningfully close analogues of other bridged assemblies such as 2, 3, and the P-cluster 34 be synthesized with stability sufficient to allow the development of function? As noted, sulfide-bridged versions of 3 have been prepared (19), but their unsupported bridges render them insufficiently stable for reactivity studies. One synthetic approach to 2 involves a Ni(μ_2 -SR)₂Fe bridge between chelated Ni(II) and an [Fe₄S₄]²⁺ cluster (34). These bridged assemblies are prepared by the techniques of small-molecule coordination chemistry and lack several desirable properties, including water solubility. It is apparent that this synthetic protocol is likely to be of limited value in obtaining stable, soluble, and structurally adjustable analogues of bridged biological assemblies. The next generation of significant

analogues most likely will utilize synthetic peptides alone or in combination with α -helical bundles as structural props, engineered so as to optimize the formation, spatial arrangement, and stability of the desired assembly. The means of peptide synthesis and structural analysis are already available. The formation of heme and Fe₄S₄-heme maquettes (35) is a clear demonstration that the peptide binding and scaffolding concept is viable, thus potentiating its application and extension to different types of complex assemblies such as, *inter alia*, the active sites of sulfite reductase, carbon monoxide dehydrogenase, heme-copper oxidases, [NiFe]-hydrogenase, and the clusters of nitrogenase.

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REFERENCES

- 1. R. Cammack, Adv. Inorg. Chem. 38, 281 (1992).
- 2. H. Beinert, R. H. Holm, and E. Münck, Science, in press.
- 3. R. H. Holm, P. Kennepohl, and E. I. Solomon, Chem. Rev. 96, 2239 (1996).
- 4. (a) D. H. Flint and R. M. Allen, *Chem. Rev.* **96**, 2315 (1996). (b) H. Beinert, M. C. Kennedy, and D. C. Stout, *Chem. Rev.* **96**, 2335 (1996).
- 5. R. H. Holm, Pure Appl. Chem. 67, 217 (1995).
- (a) S. W. Ragsdale and M.Kumar, Chem. Rev. 96, 2515 (1996).
 (b) Z. Hu, N. J. Spangler, M. E. Anderson, J. Xia, P. W. Ludden, and E. Münck, J. Am. Chem. Soc. 118, 830 (1996).
- 7. B. R. Crane, L. M. Siegel, and E. D. Getzoff, Science 270, 59 (1995).
- 8. A. Volbeda, M. H. Charon, C. Piras, E. C. Hatchikian, M. Frey, and J. C. Fontecilla-Camps, Nature 373, 580 (1995).
- 9. R. H. Holm, Adv. Inorg. Chem. 38, 1 (1992).
- 10. R. H. Holm, S. Ciurli, and J. A. Weigel, Prog. Inorg. Chem. 38, 1 (1990).
- (a) T. D. P. Stack and R. H. Holm, J. Am. Chem. Soc. 110, 2484 (1988).
 (b) T. D. P. Stack, J. A. Weigel, and R. H. Holm, Inorg. Chem. 29, 3745 (1990).
- 12. M. A. Whitener, G. Peng, and R. H. Holm, Inorg. Chem. 30, 2411 (1991).
- 13. C. Zhou and R. H. Holm, Inorg. Chem., 36, in press (1997).
- 14. J. A. Weigel and R. H. Holm, J. Am. Chem. Soc. 113, 4184 (1991).
- S. Ciurli, M. Carrié, J. A. Weigel, M. J. Carney, T. D. P. Stack, G. C. Papaefthymiou, and R. H. Holm, J. Am. Chem. Soc. 112, 2654 (1990).
- 16. J. A. Weigel, K. K. P. Srivastava, E. P. Day, E. Münck, and R. H. Holm, J. Am. Chem. Soc. 112, 8015 (1990).
- 17. T. D. P. Stack, M. J. Carney, and R. H. Holm, J. Am. Chem. Soc. 111, 1670 (1989).
- 18. J. Huang, S. Mukerjee, B. M. Segal, H. Akashi, J. Zhou, and R. H. Holm, J. Am. Chem. Soc. 119, in press (1997).
- 19. (a) L. Cai and R. H. Holm, J. Am. Chem. Soc. 116, 7177 (1994). (b) C. Zhou, L. Cai, and R. H. Holm, Inorg. Chem. 35, 2767 (1996).
- 20. J. Zhou, Z. Hu, E. Münck, and R. H. Holm, J. Am. Chem. Soc. 118, 1966 (1996).
- 21. J. Zhou, J. W. Raebiger, and C. A. Crawford, J. Am. Chem. Soc. 119, in press (1997).
- 22. L. M. Roberts and P. A. Lindahl, J. Am. Chem. Soc. 117, 2565 (1995).
- 23. J. W. Raebiger, C. A. Crawford, J. Zhou, and R. H. Holm, Inorg. Chem. 36, 994 (1997).
- (a) J. N. Butt, F. A. Armstrong, J. Breton, S. J. George, A. J. Thomson, and E. C. Hatchikian, J. Am. Chem. Soc. 113, 6663 (1991).
 (b) M. G. Finnegan, R. C. Conover, J.-B. Park, Z. H. Zhou, M. W. W. Adams, and M. K. Johnson, Inorg. Chem. 34, 5358 (1995).
- K. K. P. Srivastava, K. K. Surerus, R. C. Conover, M. K. Johnson, J.-B. Park, M. W. W. Adams, and E. Münck, *Inorg. Chem.* 32, 927 (1993).
- 26. J. Zhou, M. J. Scott, Z. Hu, G. Peng, E. Münck, and R. H. Holm, J. Am. Chem. Soc. 114, 10843 (1992).
- 27. J. B. Howard and D. C. Rees, Chem. Rev. 96, 2965 (1996).
- 28. (a) M. K. Chan, J. Kim, and D. C. Rees, *Science* 260, 792 (1993). (b) J. W. Peters, M. H. B. Stowell, S. M. Soltis, M. G. Finnegan, M. K. Johnson, and D. C. Rees, *Biochemistry* 36, 1181 (1997).
- 29. P. R. Challen, S.-M. Koo, W. R. Dunham, and D. Coucouvanis, J. Am. Chem. Soc. 112, 2455 (1990).
- 30. W. Cen, F. M. MacDonnell, M. J. Scott, and R. H. Holm, Inorg. Chem. 33, 5809 (1994).
- 31. R. E. Palermo, R. Singh, J. K. Bashkin, and R. H. Holm, J. Am. Chem. Soc. 106, 2600 (1984).
- 32. K. D. Demadis and D. Coucouvanis, Inorg. Chem. 34, 436 (1995).
- 33. S. W. Muchmore, R. F. Jack, and D. R. Dean. In *Mechanisms of Metallocenter Assembly* (R. P. Hausinger, G. L. Eichhorn, and L. G. Marzilli, eds.), chap. 6. VCH Publishers, Inc., New York (1996).
- 34. F. Osterloh, W. Saak, D. Hasse, and S. Pohl, J. Chem. Soc., Chem. Commun. 777 (1996).
- (a) D. E. Robertson, R. S. Farid, C. C. Moser, J. L. Urbauer, S. E. Mulholland, R. Pidikiti, J. D. Lear, A. J. Wand, W. F. DeGrado, and P. L. Dutton, Nature 368, 425 (1994).
 (b) B. R. Gibney, S. E. Mulholland, F. Rabanal, and P. L. Dutton, Proc. Natl. Acad. Sci. USA 93, 15041 (1996).
 (c) B. R. Gibney, F. Rabanal, J. J. Skalicky, A. J. Wand, and P. L. Dutton, J. Am. Chem. Soc. 119, 2323 (1997).